### **Research** Article



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# Effects of soy peptides on IL-1β-induced matrix-degrading enzymes in human articular chondrocytes

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#### Abstract

Background: In osteoarthritis (OA) of the most common joint disease, inflammation is deeply involved in the cartilage degradation. Specifically, inflammatory cytokines like IL-1 $\beta$  are known to enhance production of matrix-degrading enzymes like matrix metalloproteinase (MMP) and a disintegrin and metalloproteinase with thrombospondin motifs (ADAMTS). Recently, soy peptides, generated by hydrolytic enzymatic digestion of soy proteins, have been found to show anti-inflammatory activity. Thereby we investigated whether soy peptides affected IL-1 $\beta$ -induced expression of matrix-degrading enzymes in human articular chondrocytes. Methods: Articular chondrocytes derived from 4 patients with bone fracture and 11 patients with OA were cultured with 0, 10, or 100  $\mu$ M soy peptides for 4 hours. Then, the cells were stimulated with IL-1 $\beta$  for 24 hours. Non-stimulated cells were used as a control. RNA extracted from the cells was subjected to quantitative real time-PCR to assess amounts of mRNA for MMP-3 and ADAMTS-4. Results: Treatment with 100  $\mu$ M soy peptides significantly suppressed mRNA levels of MMP-3 and ADAMTS-4. Results: Treatment with 100  $\mu$ M soy peptides did not show significant suppression (p=0.91 and p=0.20, respectively). In the 100  $\mu$ M soy peptides treatment, the suppression of MMP-3 mRNA levels was correlated with that of ADAMTS-4 mRNA levels (R<sup>2</sup>=0.534). Conclusion: Soy peptides suppressed the IL-1 $\beta$ -induced expression of matrix-degrading enzymes in human articular chondrocytes. Soy peptides may have potential to ameliorate cartilage degradation in OA.

#### Introduction

Osteoarthritis (OA), the most common joint disease, is histologically characterized by degeneration of cartilage and formation of osteophytes, which lead to destruction of joints [1]. Many patients with OA suffer from pain and disability [1]. Thus, to suppress the symptoms of OA and prevent the progress of OA, novel agents should be developed.

Articular chondrocytes maintain the cartilage matrix by producing and degrading matrix proteins such as type II collagen and proteoglycans [2]. In OA, imbalance between production and degradation of the matrix protein leads to cartilage degradation [1]. Inflammation is deeply involved in the cartilage degradation in OA [1]. Inflammatory cytokines like IL-1 $\beta$  are known to be increased in synovial fluid of patients with OA [3]. In particular, IL-1 $\beta$  was reported to enhance secretion of matrix metalloproteinase (MMP)-1, -3, and -13 from chondrocytes that digest multiple types of collagens [4]. Furthermore, IL-1 $\beta$  was reported to enhance secretion of a disintegrin and metalloproteinase with thrombospondin motifs (ADAMTS)-4 and -5 from chondrocytes that digest aggrecan of a representative proteoglycan [5].

Soybeans contain a large amount of nutrients such as proteins, fats, and carbohydrates. Recently, novel biological roles of soybeans have been highlighted in addition to the roles as nutrients. Specifically, socalled soy peptides, generated by hydrolytic enzymatic digestion of soy proteins, have been reported to show various functions beneficial to health such as cholesterol-lowering properties, cognitive impairmentprevention, antiviral effects, and anti-inflammatory effects [6-12]. As to the anti-inflammatory effects, soy peptides were reported to suppress lipopolysaccharide (LPS)-induced production of cyclooxygenase-2 and prostaglandin E2 in a murine macrophage cell line [10]. Furthermore, soy peptides were reported to suppress LPS-induced production of pro-inflammatory cytokines such as IL-6 and IL-8 in human umbilical vein endothelial cells [11]. In vivo, soy-derived di- and tri-rich peptides were reported to alleviate colon and ileum inflammation in pigs with dextran sodium sulfate-induced colitis [12]. Therefore, soy peptides are expected to show anti-inflammatory effects also in human articular chondrocytes, which may lead to suppression of cartilage degradation in OA. However, effects of soy peptides on human articular chondrocytes have not been investigated to our knowledge. We here investigated whether soy peptides suppressed IL-1β-induced expression of matrixdegrading enzymes in human articular chondrocytes.

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**Key words:** A disintegrin and metalloproteinase with thrombospondin motifs-4 (ADAMTS-4), Chondrocytes, IL-1 $\beta$ , Matrix metalloproteinase-3 (MMP-3), Osteoarthritis; Soy peptides

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#### Materials and methods

#### Clinical samples and preparation of chondrocytes

Human articular cartilage samples were obtained from 4 patients with bone fracture (Normal, N1-4, 1 male; age 58 years, 3 female; mean age 78.0 years [range58-88]) at therapeutic surgery of total hip arthroplasty. Cartilage samples were also obtained from 11 patients with OA (OA1-11, 1 male; age 82 years, 10 female; mean age 78.7 years [range 66-86 years]) at therapeutic surgery of total knee arthroplasty.

The diagnosis of OA was made according to the Kellgren and Lawrence criteria [13]. Written informed consent was obtained from each of the patients and this study protocol was approved by the ethics committee of St. Marianna University School of Medicine. The study was performed in compliance with the World Medical Association Declaration of Helsinki.

For chondrocyte preparation, after careful removal of synovial tissue from cartilage samples, the cartilage was minced, washed, and treated with collagenase (Wako Pure Chemical Industries, Osaka, Japan). Isolated chondrocytes (Passage 0, P0) were washed and grown in monolayer culture in Dulbecco's modified Eagle's medium (DMEM, GIBCO, Carlsbad, CA) supplemented with 10% fetal calf serum (Wako), 100 units/ml penicillin, and 100 µg/ml streptomycin (Sigma-Aldrich, St. Louis, MO, USA) on type I collagen-coated culture dishes.

#### Treatment of chondrocytes with IL-1ß and soy peptides

Soy peptides (Hinute-AM, FUJI OIL Co. Ltd., Osaka, Japan), which were mainly composed of di- and tri-peptides, were used [14].

The prepared chondrocytes (P1-P3,  $1.0x10^7$  cells/ $\phi100mm$  dish) were cultured with 0, 10, and 100  $\mu M$  soy peptides for 4 hours. Then, the cells were treated with 1  $\mu g/ml$  IL-1 $\beta$  alone (R&D, Minneapolis, MN, USA), 1  $\mu g/ml$  IL-1 $\beta$  and 10  $\mu M$  soy peptides, and 1  $\mu g/ml$  IL-1 $\beta$  and 100  $\mu M$  soy peptides for 24 hours. Non-treated cells were used as a control.

#### Evaluation of cell viability

Cell viability was evaluated by trypan blue staining. Chondrocytes, cultured with 0, 10, or 100  $\mu$ M soy peptides for 24 hours, were harvested by treatment with 0.1% trypsin-EDTA (Sigma-Aldrich). The cell pellet, collected by centrifugation for 10 min at 1000×g, was stained with 0.5% trypan blue (Sigma-Aldrich). The number of the blue -stained (dead) cells and the total number of cells were counted using a haemocytometer under a microscopy.

#### Extraction and reverse transcription (RT) of RNA

Extraction of RNA from the chondrocyte samples and reversetranscription of the RNA samples were performed using RNeasy\* (Qiagen, Venlo, the Netherlands) and High Capacity cDNA Reverse Transcription Kits\* (Life Technologies, Rockville, MD, USA), respectively.

#### Quantitative real time PCR

Real time PCR was performed using ABI Prism 7000 Sequence Detection System (Applied Biosystems, Foster city, CA, USA), according to the manufacturer's instructions. To measure mRNA levels for MMP-3 and ADAMTS-4, 1 µg of total RNA-derived cDNA was mixed with TaqMan<sup>®</sup> Gene Expression Assays (Hs00968305\_m1 and Hs00192708\_m1, respectively, Applied Biosystems) and TaqMan<sup>®</sup> Gene Expression Master Mix (Applied Biosystems). Then the mixture was subjected to real time PCR. The thermal cycle condition was as follows: 95°C for 10 min, followed by 40 cycles of 95°C for 15 s and 60°C for 60 s. To measure mRNA levels for GAPDH, 1  $\mu$ g of total RNA-derived cDNA was mixed with 300 nM each of the forward and reverse primers and Power SYBR\* Master Mix (Applied Biosystems). Nucleotide sequences of the primers for amplification of a GAPDH DNA fragment are as follows: for GAPDH: 5'-TGGTATGGTGGAAGGACTCA and 5'-ATGCCAGTGAGCTTCCCGTT. Real time PCR was performed under the same condition as above.

#### Statistical analysis

Statistical analysis was performed by IBM SPSS Statistics (Ver.22.0). Effects of soy peptides on IL-1 $\beta$ -induced matrix-degrading enzymes were analyzed by Wilcoxon signed-rank test (p<0.05).

#### Results

## Effects of soy peptides on IL-1β-induced matrix-degrading enzymes in human articular chondrocytes

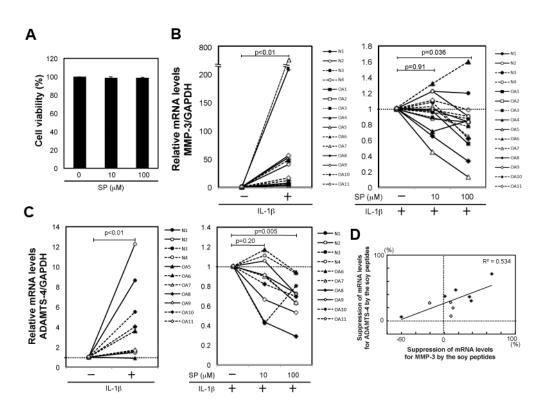
Firstly, we tested whether the concentrations of soy peptides used here showed cytotoxicity against human articular chondrocytes. As a result, we confirmed that up to 100  $\mu$ M soy peptides did not affect viability of the chondrocytes (Figure 1A). We then investigated effects of the soy peptides on IL-1 $\beta$ -induced expression of MMP-3 and ADAMTS-4 in the chondrocytes. We cultured chondrocytes with or without the soy peptides for 4 hours and then stimulated with IL-1 $\beta$ . We then assessed mRNA levels of MMP-3 and ADAMTS-4.

In the assessment of mRNA levels of MMP-3, we first confirmed that mRNA levels of MMP-3 were significantly increased by the IL-1 $\beta$  stimulation (p<0.01, Table 1 and Figure 1B, left panel). Of note, in each of the stimulated chondrocyte sample, the mRNA levels of MMP-3 were increased by the IL-1 $\beta$  stimulation. We then evaluated effects of the soy peptides on the IL-1 $\beta$ -induced expression of MMP-3 in chondrocytes. As a result, the treatment with 100  $\mu$ M soy peptides significantly

Table 1. Effects of soy peptides on IL-1 $\beta$ -induced MMP-3 mRNA levels in human chondrocytes.

Sample no.	Control	Π-1β	IL-1β +10μM HN-AM	IL-1β +100μM HN- AM
N1	0.063	13.2	16.2	15.9
N2	1.49	60.0	73.3	54.4
N3	2.13	13.5	13.0	11.9
N4	1.45	10.3	10.1	9.27
OA1	1.09	4.62	4.49	2.58
OA2	2.93	7.98	6.98	6.62
OA3	0.77	3.17	2.85	2.48
OA4	0.91	6.50	4.60	5.52
OA5	0.81	42.5	19.2	5.72
OA6	0.33	15.9	20.7	25.0
OA7	0.02	15.9	16.3	10.3
OA8	1.58	12.1	7.93	4.04
OA9	0.20	10.9	10.9	9.13
OA10	1.02	13.0	14.2	8.11
OA11	0.22	3.55	3.91	3.50
Average	1.00	15.5	15.0	11.6
STDEV	0.82	15.4	17.1	13.2
p value	-	<0.01 (vs Control)	0.91 (vs IL-1β)	0.036 (vs IL-1β)

SP: soy peptides



**Figure 1.** Effects of soy peptides on IL-1β-induced expression of MMP-3 and ADAMTS-4 in human articular chondrocytes. **(A)** Articular chondrocytes from 3 patients with OA (OA1-3) were cultured with 0, 10, and 100 µM soy peptides for 24 hours. The cells were subjected to trypan blue staining. The cell viability (%) was calculated using the following formula; (the number of unstained cells/the total number of cells)×100. SP, soy peptides. **(B)** Articular chondrocytes from 4 patients with bone fracture (N1-4) and 11 patients with OA (OA1-11) were cultured with 0, 10, and 100 µM soy peptides for 4 hours. Then, the cells were stimulated with IL-1β for 24 hours. Non-stimulated cells were used as a control. The mRNA levels for MMP-3 and GAPDH were assessed by RT-qPCR. The mRNA levels of MMP-3 were corrected using those of GAPDH. Left: The corrected mRNA level of MMP-3 was defined as 1.0 in each of the IL-1β-treated samples. Right: The corrected mRNA level of MMP-3 was defined as 1.0 in each of the IL-1β-treated samples. P-values were calculated by Wilcoxon signed-rank test using the values listed in Table 1. **(C)** Articular chondrocytes from 4 patients with bone fracture (N1-4) and 7 patients with OA (OA5-11) were treated samples. Right: The corrected mRNA level of ADAMTS-4 was defined as 1.0 in each of the IL-1β-treated samples. P-values were calculated by Wilcoxon signed-rank test using the values listed in Table 1. **(i)** articular chondrocytes from 4 patients. P-values were calculated by Wilcoxon signed-rank test using the values listed in Table 2. SP, soy peptides. **(D)** Suppression rates (%) of mRNA levels for MMP-3 and ADAMTS-4 in each of the chondrocyte samples (4 patients with bone fracture (N1-4) and 6 patients with OA (OA6-11)) were calculated by using the values listed in Table 1 and 2, respectively. A correlation diagram between the suppression of mRNA levels for MMP-3 and ADAMTS-4 in the treatment with 100 µM soy peptides treatment is shown. Coefficient of determination (R<sup>2</sup>) was calculated by regressi

suppressed mRNA levels of MMP-3 enhanced by the IL-1 $\beta$  stimulation (p=0.036, Table 1 and Figure 1B, right panel), although the treatment with 10  $\mu$ M soy peptides did not show significant suppression (p=0.91, Table 1 and Figure 1B, right panel).

In the assessment of ADAMTS-4, we first confirmed that mRNA levels of ADAMTS-4 were significantly increased by IL-1 $\beta$  stimulation (p<0.01, Table 2 and Figure 1C, left panel). Ten out of the 11 chondrocyte samples up-regulated the mRNA levels of ADAMTS-4 by the IL-1 $\beta$  stimulation. We thus evaluated effects of the soy peptides on the IL-1 $\beta$ -induced expression of ADAMTS-4 using the 10 chondrocyte samples that positively responded to IL-1 $\beta$ . As a result, the treatment with 100  $\mu$ M soy peptides significantly suppressed mRNA levels of ADAMTS-4 enhanced by the IL-1 $\beta$  stimulation (p=0.005, Table 2 and Figure 1C, right panel), although the treatment with 10  $\mu$ M soy peptides did not show significant suppression (p=0.20, Table 2 and Figure 1C, right panel).

Finally, we investigated correlation of the suppressing effect of the soy peptides on the MMP-3 expression to that of ADAMTS-4 expression. As a result, in the treatment with 100  $\mu$ M soy peptides, the suppression of MMP-3 mRNA levels was correlated with that of ADAMTS-4 mRNA levels (R<sup>2</sup>=0.534, Figure 1D). In addition, the suppressive effect of the soy peptides on the expression of MMP-3 and

ADAMTS-4 tended to be higher in patients with OA than in those with bone fracture (Figure 1D).

#### Discussion

We here found that soy peptides suppressed IL-1 $\beta$ -induced production of MMP-3 and ADAMTS-4 in human articular chondrocytes.

The possible mechanisms for the actions of soy peptides are as follows: 1) soy peptides inhibit the input of IL-1 $\beta$  signal transduction by binding to IL-1 $\beta$  or IL-1 $\beta$  receptors, 2) soy peptides, taken up into chondrocytes, inhibit the intracellular IL-1 $\beta$  signal transduction, and 3) soy peptides bind to their unknown receptors and then block the IL-1 $\beta$  signal transduction.

On the first possible mechanism, IL-1 $\beta$  activates the signal transduction system by specific binding to IL-1 $\beta$  receptors. In the majority of the tested chondrocytes samples, the mRNA levels of MMP-3 of the IL-1 $\beta$ -stimulated chondrocytes group were suppressed by the treatment with 100  $\mu$ M soy peptides (Figure 1B). However, in a small part of the tested chondrocytes samples, the mRNA levels of MMP-3 were increased by the soy peptides treatment (Figure 1B). If soy peptides bound to IL-1 $\beta$  or IL-1 $\beta$  receptors, the mRNA levels of MMP-

Sample no.	Control	IL-1β	IL-1β +10μM HN-AM	IL-1β +100μM HN-AM
N1	0.16	1.37	1.25	1.00
N2	0.23	2.78	2.94	2.00
N3	0.13	0.74	0.31	0.59
N4	0.92	1.49	1.65	1.38
OA5	2.03	1.77	-	-
OA6	1.30	4.71	5.53	4.43
OA7	1.70	2.96	2.66	1.88
OA8	1.02	1.51	0.66	0.43
OA9	1.47	2.20	1.46	1.16
OA10	0.91	3.64	2.98	2.52
OA11	1.11	1.83	1.66	1.14
Average	1.00	2.27	2.11	1.65
STDEV	0.63	1.16	1.50	1.17
p value	-	<0.01 (vs. Control)	0.20 (vs. IL-1β)	0.005 (vs. IL-1β)

Table 2. Effects of soy peptides on IL-1 $\beta$ -induced ADAMTS-4 mRNA levels in human chondrocytes.

SP: soy peptides

3 would have been suppressed in all the tested chondrocytes samples. Thus, it would be unlikely that soy peptides inhibit the binding between IL-1 $\beta$  and IL-1 $\beta$  receptors.

On the second possible mechanism, we showed that suppression of mRNA levels for MMP-3 were well correlated with that of mRNA levels for ADAMTS-4 in the soy peptides-treated chondrocyte samples (R<sup>2</sup>=0.5343, Figure 1D). Thus, we thought that soy peptides suppressed the IL-1 $\beta$ -induced production of MMP-3 and ADAMTS-4 by inhibiting their common pathway. IL-1β activates the signal transduction systems through the activation of multiple protein kinases such as p38 mitogenactivated protein kinase (MAPK), c-Jun N-terminal kinase (JNK), and Akt [15-18]. The activated MAPK, JNK, and Akt enhance production of various inflammatory cytokines and matrix degrading proteins, activating transcription factors such as AP-1 and NF-KB [19]. Akt, phosphorylated by activated phosphoinositol 3-kinase (PI3K), leads to activation of NF-KB [20]. NF-KB activates transcription of proinflammatory cytokines and matrix-degrading enzymes including MMP-3 and ADAMTS-4 [4,5]. Peptides derived from β-conglycinin of a representative soy protein have been reported to inactivate NF-kB by suppressing phosphorylation of Akt in liver and aorta of LPS-induced inflammatory model mice [11]. Thus, we speculate that soy peptides suppress the IL-1β-induced production of MMP-3 and ADAMTS-4, probably by inhibiting their PI3K-Akt-NF-κB signaling by suppressing the phosphorylation of Akt.

On the third possible mechanism, although receptors for soy peptides have not yet been identified, such receptors may exist and deliver signals. For example, M. Chimen *et al.* reported that PEPITEM, a peptide proteolytically derived from 14.3.3 zeta delta protein, used cadherin-15 as a receptor and lead to release of sphingosine-1-phosphate from endothelial cells [21]. Like the case of PEPITEM, receptors for soy peptides may exist, which should be investigated. Including these 3 possible mechanisms, the mechanisms for the action of soy peptides should be investigated in the near future.

We here found that soy peptides significantly suppressed the IL-1 $\beta$ -induced production of MMP-3 and ADAMTS-4 in human articular chondrocytes. In the majority of the tested chondrocytes samples, both the mRNA levels of MMP-3 and those of ADAMTS-4 were suppressed by the treatment with 100  $\mu$ M soy peptides (Figure 1C). However, in a small part of the tested chondrocytes samples, neither the mRNA levels of MMP-3 nor those of ADAMTS-4 were suppressed by the soy peptides treatment (Figure 1C). Thus, the effect of soy peptides would be influenced by the individual genetic backgrounds. There might be subpopulations that show different sensitivity to soy peptides.

It is interesting that the suppressive effect of the soy peptides on the expression of MMP-3 and ADAMTS-4 tended to be higher in the patients with OA than in those with bone fracture (Figure 1D). OA chondrocytes might show higher sensitivity to soy peptides than normal chondrocytes.

Soy peptides used in this study are mainly composed of di- and tri-peptides [14]. However, specific peptides that suppressed the IL-1 $\beta$ -induced production of MMP-3 and ADAMTS-4 have not been identified yet. Recently, a tri-peptide of Val-Pro-Tyr, derived from glycinin of a major soy protein, has been reported to down-regulate the expression of pro-inflammatory cytokines in intestinal epithelial cells and macrophages [22]. The soy peptides used in this study are expected to contain glycinin and thus would contain Val-Pro-Tyr. A tri-peptide of Val-Pro-Tyr might be one of the soy peptides that suppressed the effects of IL-1 $\beta$  in this study. On this point, further investigation is required.

Our data showed that soy peptides suppressed the IL-1 $\beta$ induced production of MMP-3 and ADAMTS-4 in human articular chondrocytes *in vitro*. Our data indicate that soy peptides may be useful to prevent or suppress cartilage degradation in OA.

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#### Conflict of interest statement

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