

# Mineral transfer between enamel and artificial saliva

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## Abstract

**Objectives:** The objective of this study was to clarify changes in enamel mineral density after immersion in pH-regulated artificial saliva as well as concentrations of calcium (Ca) and phosphorous (P) in artificial saliva.

**Methods:** Three segments of size 5-by-5 mm taken from the enamel surface of each of 15 selected bovine anterior teeth were prepared and coated with nail varnish, excluding the labial side. The specimens were immersed in 10 mL of artificial saliva (0.7 mM/L CaCl<sub>2</sub>, 0.2 mM/L MgCl<sub>2</sub>, 4.0 mM/L KH<sub>2</sub>PO<sub>4</sub>, 30.0 mM/L KCl, 20.0 mM/L HEPES) and regulated at pH 7.0 or 5.5 by KOH for 10 days. Their average Hounsfield unit values (HUV) were measured from the enamel surface to a site 600  $\mu$ m in depth by using Micro CT. Further, concentrations of Ca and P in the artificial saliva before and after the immersion were measured. Subsequently, the specimens of the pH 5.5 group were again immersed in artificial saliva regulated at pH 7.0 for 10 days and the same measurements were carried out.

**Results:** Average HUVs of the pH 5.5 group showed significantly lower values than the ST group only at a layer 0-150  $\mu$ m in depth ( $p<0.01$ ). The concentrations of Ca and P in the pH 5.5 group showed significantly higher values after the immersion ( $3.73 \pm 0.44$ ,  $12.37 \pm 0.41$ ) than those before the immersion ( $2.80 \pm 0.18$ ,  $11.72 \pm 0.12$ ). As a result of immersion of specimens of pH 5.5 group in saliva of pH 7.0, there was no significant change in mineral density in enamel and Ca and P in saliva.

**Conclusion:** It was confirmed that the mineral transfer between enamel and saliva in demineralization depends on the pH of saliva.

## Introduction

It is possible to confirm mineral transfer between enamel and saliva by analyzing changes in mineral density in saliva [1]. On the other hand, few studies have been made on the aspect of increase and decrease in minerals on the enamel side, leaving the details unknown.

Even though the Transversal Micro Radiography (TMR) method [2] is generally used for evaluating tooth demineralization and remineralization by observing teeth segments using an X-ray Micro Analyzer, it is impossible to observe continuous change in demineralization due to limitation in preparation of specimens by this method. The Quantitative Light Induced Fluorescence (QLF) method is applicable for clinical evaluation for demineralization and remineralization in a non-destructive manner, but it is possible neither to measure a slight amount of demineralization for a short period of time nor to observe it by each depth [3]. On the contrary, Micro X-ray CT (Micro CT) with a property capable of sterically observing change in mineral density in small amounts by micro units has been used recently for measuring change in enamel mineral density [4] during early stage demineralization or enamel mineral density with developed dysplasia [5].

The objective of the study was to analyze changes in the concentration of Ca and P in artificial saliva and variation in mineral density on the enamel surface layer based on Micro CT using bovine teeth enamel and artificial saliva with regulated pH in order to clarify the actual conditions of mineral transfer between mineral and saliva.

## Materials and methods

### Materials

Polishing 15 bovine anterior teeth with a sandpaper (No. 400-600) moderately and taking tomographic images to be reconstructed

by using Micro CT (SKYSCAN1172, SKYSCAN, Kontich, Belgium) with a tube voltage of 100 kVp and tube current of 100  $\mu$ A, the average Hounsfield unit value (HUV) of enamel from the surface to a site 600  $\mu$ m in depth was measured [6]. In order to exclude teeth with larger inter-individual variability in HUV, a total of 10 of them were served for the experiment by eliminating 5 teeth with an SD exceeding the average value of 1.

### Methods

**Analysis of specimens:** Preparing three pieces of tooth tissue specimens, with a size of 5x5x5 mm including enamel surface on the labial side, from each selected bovine tooth, each specimen was immersed in 10 mL of artificial saliva separately within an incubator which was kept at 37  $^{\circ}$ C for 10 days after coating all surfaces other than the labial side of each specimen with nail varnish. Extracting all specimens from the artificial saliva 10 days after the immersion and removing the nail varnish with polish remover, tomographic images taken by using Micro CT under a condition of a tube voltage of 100 kVp and tube current of 100  $\mu$ A was reconstructed and then, the average

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**Special Issue: Early Caries (Surfaces Demineralization)**

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HUV of enamel from the surface at a site 600  $\mu$ m in depth was measured (Figure 1). In the subsequent remineralization experiment, the average HUV was measured similarly after immersing in remineralization solution regulated at pH 7.0.

**Artificial salivary constituent:** Artificial saliva was prepared by a formulation (0.7 mM of  $\text{CaCl}_2$ , 0.2 mM of  $\text{MgCl}_2$ , 4.0 mM of  $\text{KH}_2\text{PO}_4$ , 30.0 mM of  $\text{KCl}$ , and 20.0 mM of HEPES) [7] commonly used for enamel remineralization experiments or the like while adjusting at pH 7.0 or 5.5 by potassium hydroxide. Specimens immersed in artificial saliva at pH 7.0 and 5.5 were classified into a pH 7.0 group and pH 5.5 group respectively, with the control (ST) group which were immersed in neither pH of artificial saliva. Concentrations of Ca and P were measured just before immersing specimens as well as soon after 10 days of immersion. In the remineralization experiment, specimens immersed in artificial saliva at pH 5.5 for 10 days were further immersed in 10mL of artificial saliva at pH 7.0 which was replaced by new saliva every two days, and concentrations of Ca and P were measured each time saliva was replaced. In the case of the ST group, four specimens of bovine anterior teeth enamel, of which HUVs had been measured by Micro CT after demineralization, were further immersed in ultrapure water regulated at pH 7.0 for 5 days and their HUVs were measured.

**Measurement of Ca concentration in artificial saliva:** A calcium measurement kit (Calcium E- Test Wako/Wako Pure Chemical Industries Ltd., Japan) was used for measurement of Ca concentration. Consisting of a buffer solution (monoethanolamine buffer solution/pH 12.0), coloring test solution (methyl xylene blue MXB, 8-quinolinol) and Ca standard solution, the Calcium E – Test Wako utilizes a reaction which appears to be blue by combining Ca with MXB under alkaline

conditions. Preparing a calibration curve by using an Absorbance Microplate Reader, the Ca concentration was obtained based on absorbance of each sample.

**Measurement of P concentration in artificial saliva:** A phosphorous measurement kit (Phosphor C- Test Wako/Wako Pure Chemical Industries Ltd., Japan) was used for measurement of Phosphorous concentration. Consisting of a coloring test solution (p-methylamino phenol sulfate, surfactant, surfactant, ammonium molybdate, sulfuric acid), and P standard solution, Phosphor C-Test Wako utilizes a reaction in which phosphomolybdc acid generated by combining P with molybdate appears to be blue in a form of molybdenum blue further reduced by p-methylamino phenol sulfate. Preparing a calibration curve by using an Absorbance Microplate Reader similarly to the case of Ca concentration, P concentration was obtained based on the absorbance of each sample.

**Statistical processing:** In terms of the comparison of measurement results, a Student's t-test was used for comparisons between two groups, and the 2-way ANOVA as well as Scheffe's method were used for those between more than two groups, regarding values with  $p < 0.05$  to be significant.

## Results

Changes in Ca and P concentrations before and after immersion of the specimens in artificial saliva at each pH for 10 days are shown in Table 1. Ca concentration in saliva regulated at pH 7.0 was  $1.73 \pm 0.44$  mg/dL and  $1.51 \pm 0.13$  mg/dL before and after the immersion respectively, showing no significant difference between the two values. On the other hand, that in saliva regulated at pH 5.5 was  $2.80 \pm 0.18$  mg/

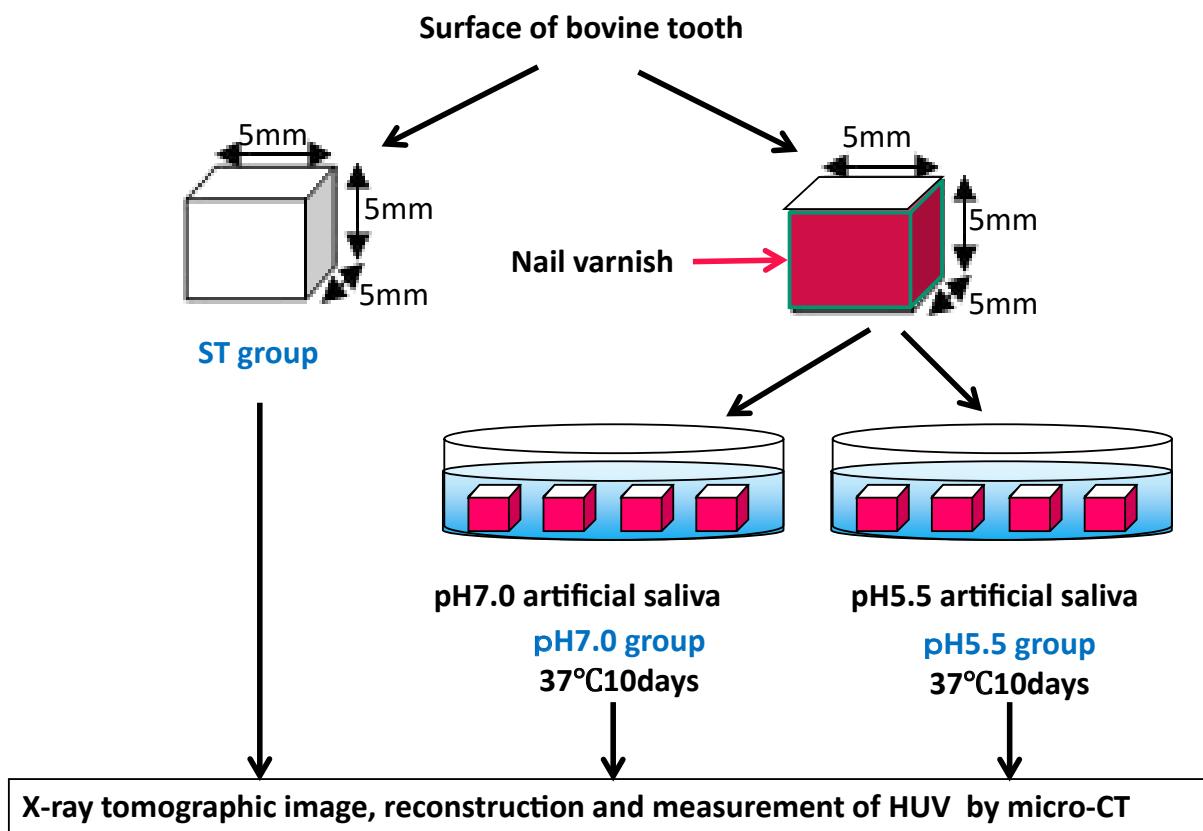


Figure 1. Method

dL and  $3.73 \pm 0.44$  mg/dL before and after the immersion respectively, with significant increase after immersion recognized ( $p<0.05$ ).

No significant difference was observed in the values of P concentration between before and after immersion in saliva regulated at pH 7.0. In the case of saliva regulated at pH 5.5, the value after immersion ( $12.37 \pm 0.41$  mg/dL) showed significant increase ( $p<0.01$ ) compared with that before immersion ( $11.72 \pm 0.12$  mg/dL).

Specimens from the pH 5.5 group were immersed into artificial saliva regulated at pH 7.0, which was used previously, in order to cause remineralization (RM). Concentrations of Ca and P observed in the artificial saliva, which was replaced every two days, are shown in Table 2. As the result, all values of Ca concentration measured 5 times after two to 10 days after immersion were lower values than the value  $1.73 \pm 0.44$  mg/dL before immersion while showing no significant difference. Similarly, P concentration before immersion was  $11.68 \pm 0.15$  mg/dL showing no significant difference between values at each measurement.

Table 3 shows the comparison of values between ST, pH 7.0 and pH 5.5 groups and the average HUVs of RM for each depth. The pH 5.5 group showed significantly lower values than the ST group only at a layer 0-150 $\mu$ m in depth ( $p<0.01$ ). On the other hand, no significant difference was observed in the case of the pH 7.0 group in values of any layer compared with those of the ST group. Significant difference was observed in the case of RM in values of all layers compared with ST, pH 5.5 and pH 7.0 groups ( $p<0.01$ ).

As a control group of RM, Table 4 shows the results of HUVs measured after immersing four specimens, of which HUVs had been measured after demineralization in ultrapure water regulated at pH 7.0 for 5 days. There was no significant difference between the average HUV of each layer and values soon after demineralization (DM).

**Table 1.** Changes in the concentration of Ca and P in the artificial saliva (mg/dL) (n=10)

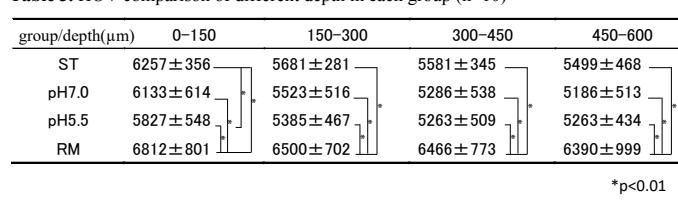
	pH7.0		pH5.5	
	Ca	P	Ca	P
Before immersion	$1.73 \pm 0.44$	$11.68 \pm 0.15$	$2.80 \pm 0.18$	$11.72 \pm 0.12$
After immersion	$1.51 \pm 0.13$	$11.77 \pm 0.13$	$3.73 \pm 0.44$	$12.37 \pm 0.41$

\*:p < 0.05      \*\*:p<0.001

**Table 2.** Concentration of Ca and P in artificial saliva (RM) during remineralization (mg/dL) (n=10)

	Before immersion	2days	4days	6days	8days	10days
Ca	$1.73 \pm 0.44$	$1.49 \pm 0.15$	$1.55 \pm 0.15$	$1.51 \pm 0.25$	$1.39 \pm 0.25$	$1.54 \pm 0.46$
P	$11.68 \pm 0.15$	$11.54 \pm 0.08$	$11.58 \pm 0.26$	$11.75 \pm 0.24$	$11.67 \pm 0.32$	$11.50 \pm 0.31$

**Table 3.** HUV comparison of different depth in each group (n=10)



\*p<0.01

**Table 4.** HUV of the control group at the time of remineralization (n=4)

group/depth(μm)	0-150	150-300	300-450	450-600
DM	$6110 \pm 619$	$5993 \pm 509$	$5909 \pm 448$	$5926 \pm 391$
RM	$6294 \pm 411$	$6058 \pm 231$	$5937 \pm 128$	$5971 \pm 270$

DM: after demineralization

## Discussion

### Changes in Ca and P concentration in artificial saliva

Mineral transport between enamel and saliva is caused by changes in mineral solubility resulting from changes in the pH of saliva [1]. More specifically, when the pH of resting saliva is a normal pH ranging from 6 to 6.5, salivary minerals are saturated with respect to hydroxyapatite and mineral migration does not occur. However, if the secretion rate of saliva increases and the pH rises [8], mineral solubility in saliva decreases and dissolved minerals precipitate or are absorbed by enamel. On the other hand, in the case of the decrease in salivary secretion rate typified by xerostomia, salivary minerals become unsaturated with respect to hydroxyapatite (solubility increases), and minerals begin to be released into saliva [9]. Fosdic, *et al.* [1] and Ericsson [10] report that the pH around pH 5.5 is the critical pH. In the present study, experiments were conducted by adjusting the pH of artificial saliva to 5.5 in order to clarify the demineralization of enamel at the critical pH. Also, as for artificial saliva, that prescribed by ten Cate [7], which has been used in many remineralization studies, was used.

The results of this study are that in the case that enamel was immersed in artificial saliva at pH 7.0, no statistically significant change was observed in the Ca and P concentrations in the artificial saliva after immersion, but that in the case of the specimen immersed in artificial saliva at pH 5.5, a statistically significant increase was observed in the Ca and P concentrations in the artificial saliva. This suggests that minerals in the artificial saliva were saturated with respect to enamel at pH 7.0, and that no statistically significant mineral migration occurred between them, whereas because at pH 5.5, minerals in the artificial saliva were unsaturated with respect to enamel, minerals were released from the enamel to the artificial saliva.

After that, when the specimen immersed in the pH 5.5 group was immersed in artificial saliva at pH 7.0, the Ca concentration in artificial saliva measured every 2 days was lower than that before immersion. In this experiment, artificial saliva was replaced every 2 days in order to provide more fresh artificial saliva to the specimen. It is inferred that if immersion had been carried out with the same artificial saliva for more than two days, a statistically significant difference would have been observed in the Ca concentration between before and after immersion.

### Changes in mineral density in enamel

Enamel contains many trace elements. Brudevold and Soremark [11] divided enamel into three layers from the surface layer to the inside, examined the concentration of trace elements by powdering each layer. They reported that F, Zn, Pb, Fe, Sn are highly concentrated in the surface layer, that Sr and Cu are almost evenly distributed throughout the three layers, and that CO<sup>2</sup> Mg and Na are more highly concentrated in the deep layer. The fact that F and Zn are abundantly present in the layer where subsurface demineralization occurs means that even if demineralization occurs, these elements are in an environment in which they can be easily absorbed in the process of remineralization when the environment improves. When F ions are substituted in apatite crystals, stable crystals of fluoroapatite,  $\text{Ca}_{10}(\text{PO}_4)_6\text{F}_2$ , are formed. It is known that the acid resistance of fluoroapatite [12] increases sharply as the F content increases.

The results of this study were that when the mineral density in the region from the surface layer down to the depth of 600  $\mu$ m was examined in the ST group, the HUV was highest on the surface and decreased as it went deeper. This indicates that besides the minerals contained in enamel, the mineral density is higher in the enamel

surface layer than in the deep layer because the enamel surface layer is constantly influenced by saliva, etc. [13]. This is the reason that clinically demineralization of enamel begins from the region under the surface layer where mineral density is lower than in the surface layer rather than from the surface layer with relatively good acid resistance.

Although we were unable to identify the minerals in this study, we were able to clarify the transition of mineral density from the surface layer to the deep layer for the first time by using micro CT.

Enamel repeats demineralization and remineralization daily due to changes in saliva pH in the oral cavity. The remineralization therapy is considered to increase its importance in dental clinic in the future. The results of the experiments in this study confirmed aspects of enamel-salivary mineral migration in initial demineralization. This seemed to be useful as basic data on the future remineralization therapy and the maturation of immature permanent teeth.

#### References

1. Fosdick LS, Starke AC (1939) Solubility of tooth enamel in saliva at various pH levels. *J Dent Res* 18:269-263.
2. Hafström-Björkman U, Sundström F, de Josselin de Jong E, Oliveby A, Angmar-Måansson B (1992) Comparison of laser fluorescence and longitudinal microradiography for quantitative assessment of in vitro enamel caries. *Caries Res* 26:241-247. [\[Crossref\]](#)
3. Ando M, van der Veen MH, Schemehorn BR, Stookey GK (2001) Comparative study to quantify demineralized enamel in deciduous and permanent teeth using laser and light-induced fluorescence techniques. *Caries Res* 35:464-470. [\[Crossref\]](#)
4. Huang TT, Jones AS, He LH, Darendeliler MA, Swain MV (2007) Characterisation of enamel white spot lesions using X-ray micro-tomography. *J Dentistry* 35:737-43. [\[Crossref\]](#)
5. Farah RA, Swain MV, Drummond BK, Cook R, Atieh M (2010) Mineral density of hypomineralised enamel. *J Dentistry* 38:50-58. [\[Crossref\]](#)
6. Wong FS, Anderson P, Fan H, Davis GR (2004) X-ray microtomographic study of mineral concentration distribution in deciduous enamel. *Arch Oral Biol* 49:937-944. [\[Crossref\]](#)
7. ten Cate JM, Duijsters PPE (1982) Alternating demineralisation and remineralization of artificial enamel lesions. *Caries Res* 16:201-210. [\[Crossref\]](#)
8. Dawes C, Dong C (1995) The flow rate and electrolyte composition of whole saliva elicited by the use of sucrose-containing and sugar-free chewing-gums. *Arch Oral Biol* 40:699-705. [\[Crossref\]](#)
9. Silverstone LM (1977) Remineralization phenomena. *Caries Res* 11: 59-84. [\[Crossref\]](#)
10. Ericsson Y (1949) Investigations into the calcium phosphate equilibrium between enamel and saliva and its relation to dental caries. *Acta Odont Scand* 8.
11. Brudevold F, Soremark R (1967) Chemistry of the mineral phase of enamel from structural and chemical organization of teeth. Edited by Miles AEW, Academic Press New York and London, 247-277.
12. Poulsen S, Larsen MJ (1975) Dental caries in relation to fluoride content in the primary dentition. *Caries Res* 9:59-65. [\[Crossref\]](#)
13. ten Cate JM, Larsen MJ, Pearce EIF, Fejerskov O (2008) Fluoride reactions in the oral environment. In *Dental Caries, The Disease and its Clinical Management* (2<sup>nd</sup> edition), Ole Fejerskov and Edwina Kidd (ed), BlakwellMunksgaard, Oxford, 225-226.