

# Enhancement of clonal growth of normal human keratinocytes in a serum-free medium by Lithium ions, Dibutyl-Cyclic AMP and Prostaglandin E<sub>1</sub>

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## Abstract

A serum free medium (MCDB153) was designed to support the clonal growth of normal human keratinocytes (NHK) when supplemented with two protein growth factors, insulin-like growth factor-1 (IGF-1) and epidermal growth factor (EGF). We now report the results of new clonal growth studies showing enhanced clonal growth of NHK by three new non-protein growth factors. Newborn NHK cells were plated at 500 cells/cm<sup>2</sup> in serum-free basal media supplemented singly with one of the following additives: di-butyl cyclic AMP (diBcAMP, 1 × 10<sup>-4</sup>M), lithium chloride (10 mM), and prostaglandin E<sub>1</sub> (10 ug/ml) in media supplemented with either IGF-1 only or IGF-1 plus EGF. All dishes were fixed ten days later stained with crystal violet stain (0.2%) and photographed. DiBcAMP in combination with IGF-1 enhanced clonal growth relative to control dishes supplemented with IGF-1 plus EGF, indicating that diBcAMP can completely eliminate EGF as a necessary growth factor. Likewise single addition of lithium ions to SFM in combination with IGF-1 enhanced clonal growth relative to controls supplemented with IGF-1 plus EGF. Finally, PGE<sub>1</sub> enhanced NHK clonal growth in dishes supplemented with either IGF-1 alone, with IGF-1 plus EGF and even with no protein growth supplementation relative to control dishes supplemented with IGF-1 plus EGF. PGE<sub>1</sub> appears to eliminate the need for both EGF and IGF-1. A second study was undertaken to examine the enhancing effect of non-protein factors in the presence of elevated levels (3X) of six key amino acids (his, met, phe, tryt, ileu and tyr) that allow attainment of higher clonal cell densities. Elevated amino acids actually suppressed the enhancing effect of lithium ions on clonal growth. By contrast, elevated amino acids in SFM medium supplemented with PGE<sub>1</sub> in combination with IGF-1 displayed enhanced clonal growth relative to control dishes supplemented with EGF plus IGF-1 in the elevated amino acid medium, suggesting a possible synergetic action of PGE<sub>1</sub> and elevated amino acids. These results provide new insights in to the complex interplay between, non-protein ligands, nutritional factors and protein growth factor stimulation.

## Introduction

The successful serum-free culture of normal epidermal keratinocytes was reported earlier [1-8]. Serum-free culture of NHK have a strict requirement for two protein factors, epidermal growth factor (EGF) and insulin (Ins) [9]. Insulin can be replaced with insulin-like growth factor-1 (IGF-1) [10,11]. Long-term culture of NHK often require further supplementation with bovine pituitary extracts (BPE), but can be avoided in early passage cultures. Elimination of either or both EGF and insulin/IGF-1 would be a significant advance as they are the most expensive ingredients in a defined SFM and are cost prohibitive for production of cultured epidermal autograft (CEA) production.

We have investigated the effect of several non-protein agents that appear to replace EGF and or insulin for the clonal growth of NHK. Among these are Lithium ions, dibutryl-cycluc adenosine monophosphate (diBcAMP), and prostaglandin E<sub>1</sub> (PGE<sub>1</sub>). Previous studies have documented the need for many trace element metal ions in defined basal nutrient media and are included in many present day SFM compositions cited above. Here we report that Lithium ions stimulate the multiplication of human keratinocyte cells in a serum-free chemically defined medium. Lithium ions (Li<sup>+</sup>) can substitute for sodium ions in biological systems. It has a smaller atomic radius, it forms stronger ionic bonds. Lithium affects DNA and RNA synthesis and cell multiplication of mouse mammary gland explants cultured in a chemically-defined synthetic medium [12]. We also examined the effect of dibutryl-cyclic 3'-5'-cyclic adenosine monophosphate (dB-cAMP) on the clonal growth of normal human keratinocytes in

a serum-free chemically defined culture medium either singly or in combination with Lithium ions. cAMP through cAMP-dependent PKA regulates actin organization and cell motility [13]. cAMP does not readily cross the plasma membrane of mammalian cells. Therefore, when one studies the action of cAMP on cells in culture the more membrane-soluble derivative, dibutryl-cyclic AMP (diBCAMP) is usually employed. An early study reported that diBCAMP inhibited epidermal cell division [14]. Addition of cholera toxin, a potent toxin, which induces an increase in intracellular cAMP, actually enhances keratinocyte growth [15]. Although, cholera toxin had no effect on human keratinocyte growth in a fetal serum-supplemented medium [1]. These disparate results may also reflect complex interactions and unpredictable consequences attributable to employing serum-containing culture media. cAMP is a necessary growth factor for the clonal growth of urothelial cells cultures in SFM [16].

Finally, we investigated the effect of prostaglandin E<sub>1</sub> (PGE<sub>1</sub>) on

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**Key words:** Dibutry-3',5'-cyclic adenosine monophosphate, lithium ions, prostaglandin E<sub>1</sub>, serum-free media, non-protein growth factors, insulin-like growth factor-1, epidermal growth factors, normal human keratinocytes

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NHK clonal growth. Little is known about its effects on proliferation of epidermal keratinocytes. Human skin generates eicosanoids, which have been reported to regulate growth and differentiation of the epithelia but do not play a central role in inflammatory skin diseases [17]. PGE<sub>1</sub> is a potent vasodilator and antithrombotic and injection of it have been widely used in circulatory disturbances in skin ulcers and various collagen diseases. PGE1 has shown clinical effectiveness for treatment of burn wounds [18]. PGE1 stimulates chloride secretion in a colonic epithelial cell line, which is associated with an increase in cyclic AMP level [19]. In addition, PGE<sub>1</sub> is reported to increase EGF production in three-dimensional cultured human annulus cells [20]. A combination of ovine prolactin (0.1 microgram/ml) and PGE<sub>1</sub> supports clonal growth of early passage human mammary epithelial cells in a serum-free chemically defined synthetic culture medium and has been included in selective media for keratinocytes and fibroblasts [4].

**Material and methods**

**Materials**

All chemicals were purchased from Sigma- Aldrich (St. Louis, MO). Cultures dishes were from Corning (Corning, NY). MCDB153 serum-free basal nutrient media were made fresh as previously described [11].

**Clonal growth assay/cell culture**

Methods for isolating and culturing primary and secondary-passage normal human neonatal normal human keratinocyte (NHK cells) were those previously employed [9]. Clonal growth assays were performed as previously described [9]. Secondary serially-passage cultures were seeded in duplicate sterile disposable 35 mm<sup>2</sup> Petri dishes at 500 cells/cm<sup>2</sup> in a MCDB153 SFM basal media and supplemented singly with the following additives: diBCAMP (1 × 10<sup>-4</sup> M), LiCl (10 mM), and PGE1 (10 µg/ml) with or without further supplementation with either or both insulin-like growth factor (IGF-1, 5 ng/mL) and epidermal growth factor (EGF, 5-10 ng/mL). All dishes were fixed ten days later stained with crystal violet stain (0.2%) and photographed for later analysis.

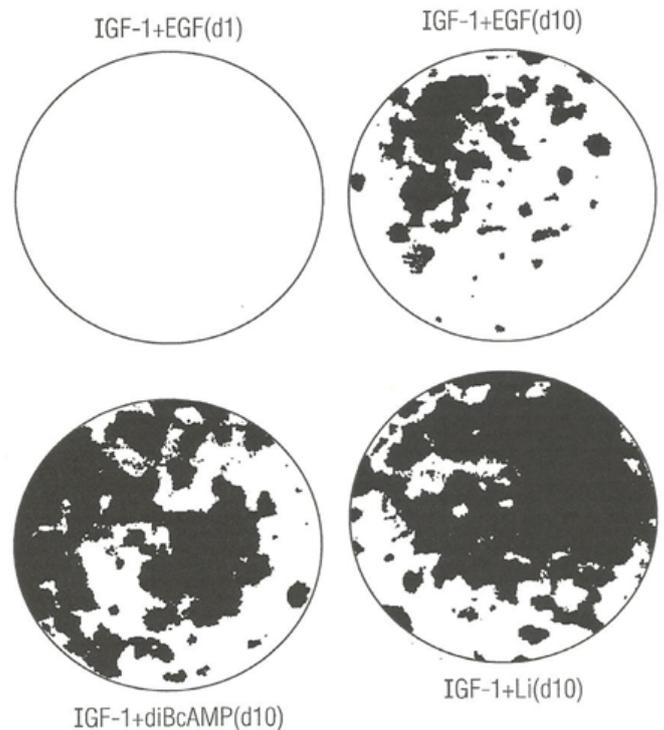
**Results**

**Growth stimulation of NHK by single addition of DiBCAMP and Li Cl**

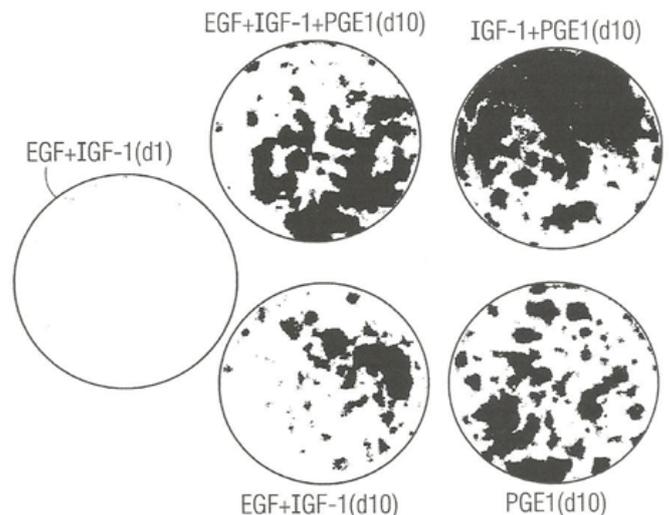
Figure 1 presents photographs showing four different clonal growth assay dishes (A-D). The effect of single addition of diBCAMP (C) or Lithium Chloride (D) to SFM supplemented only with IGF-1 and fixed and stained after day 10 of clonal growth compared with SFM supplemented with both EGF and IGF-1 and fixed and stained after day 1 (A) and after day 10 (B) of clonal growth. The results show that both dBcAMP and Lithium Chloride not only have EGF-like activity as they replace EGF but actually enhance clonal growth relative to the combination of EGF plus IGF-1. One important implication of these results is the potential benefit of replacing expensive EGF with Lithium Chloride or diBCAMP as low cost non-protein growth factors.

**Enhancement of clonal growth of NHK by single addition of Prostaglandin E1**

Figure 2 presents photographs showing 5 different clonal growth assay dishes. There is only negligible clonal growth after just 1 day of growth (A) and much greater clonal growth 10 days later (D) in control SFM dishes supplemented with EGF plus IGF-1 as expected. Addition of PGE<sub>1</sub> to SFM dishes already supplement with EGF and IGF-1 (B)



**Figure 1.** Photographs showing the enhancing effect of diButyl cyclic adenosine monophosphate (diBcAMP) and Lithium ions (Li<sup>+</sup>) on clonal growth of normal human keratinocytes. A) Control clonal growth assay dishes: (top left), labeled IGF-1+EGF, d1 (day one) and (top right), labeled IGF-1 + EGF, d10 (day ten). B) Experimental clonal growth assay dishes: culture medium containing IGF-1 (5 ng/mL) supplemented with diBcAMP (0.1 mM) (bottom left), labeled IGF-1 + diBcAMP; C) culture medium supplemented with IGF-1 (0.1 mM) and Li<sup>+</sup> ions (bottom right). All dishes were fixed with glutaraldehyde and stained with 0.2% crystal violet stain. The dishes were photographed (IX, magnification).



**Figure 2.** Photograph showing the enhancing effect of prostaglandin E1 (PGE1) in combination with IGF-1 or insulin (Ins) in the presence or absence of epidermal growth factor (EGF) on the clonal growth of normal human keratinocytes. A) Control culture dishes: (top left), labeled EGF + IGF-1, (d1, day one); (middle bottom), labeled EGF + IGF - 1 (d10, day 10). B) Experimental culture dishes: (top middle), labeled EGF + IGF-1 + PGE1, d10 (day ten); (top right), labeled IGF-1 + PGE1 (d10); bottom right- labeled PGE1 (d 10). All dishes were fixed with 5% glutaraldehyde and stained with 0.2% Crystal Violet stain and. The dishes were photographed (IX, magnification).

and fixed and stained 10 days later exhibited enhanced clonal growth relative to control dish (D). Experimental dish (C) was supplemented

with PGE1 and IGF-1 and no EGF. It greatly enhanced clonal growth relative to the control dish (D), indicating that PGE1 can replace EGF as a necessary growth factor, and exhibited greater clonal growth than mere addition of PGE1 to SFM already supplemented with EGF and IGF-1. Finally, we tested the effect of single addition of PGE1 to SFM without any protein growth factors (E). It exhibited clonal growth equal to PGE1 in combination with EGF and IGF-1, suggesting that the entire enhancement of PGE1 in combination with both EGF and IGF-1 may be due to PGE1 alone.

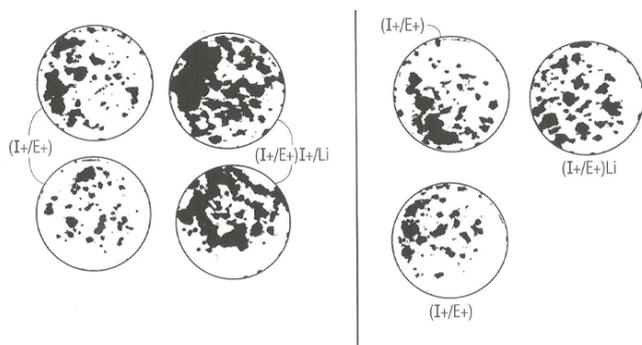
**Effect of elevated levels of key amino acid on PGE1 and Li<sup>+</sup> ion - induced enhancement of clonal growth**

In an independent study, we examined the effect of elevated 6 key amino acids (his, ileu met, phe, tryt, and tyr) at three-times their normal concentration in MCDB153 basal nutrient medium on the effect of lithium chloride and PGE1. Figure 3 (right panel) shows that mere elevation of the six key amino acids (B) had a negative stimulatory effect on clonal growth of duplicate SFM dishes ((A, C) supplemented with Lithium Chloride and EGF and IGF-1 relative to duplicate control and Lithium Chloride dishes with the 1X level of amino acids (left panel). This may indicate that Li<sup>+</sup> ion is sequestered in medium containing high levels of amino acid.

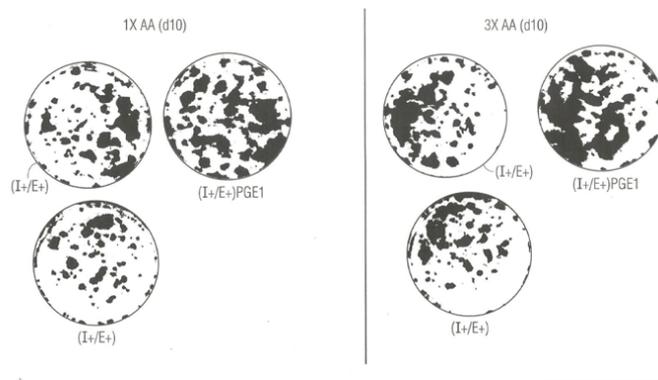
By contrast, there was a significant enhancements of clonal growth in culture medium supplemented with PGE1 containing elevated levels of the six key amino acids (right panel) relative to the lower (1X) levels of amino acid when cultured in the presence of EGF and IGF-1 protein growth factors. This result indicates that the strength of the stimulatory effect of PGE1 is dependent on the nutritional action of elevated levels of amino acids.

**Discussion**

A cost-effective serum-free culture of human epidermal keratinocytes is of paramount importance for the *in vitro* formation of cloned human tissues by means of cell therapy, and eventually for the success of regenerative medicine’s approach to replace diseased, injured, “worn-out/aged” and lost host tissue. This can be accomplished by eliminating either EGF or IGF-1 from a complete culture growth medium. Here, we have described three different non-protein growth promoting factors. Each is an additive that can be added to a chemically defined serum-free culture medium suitable for the growth and proliferation of rapidly growing normal human keratinocytes.



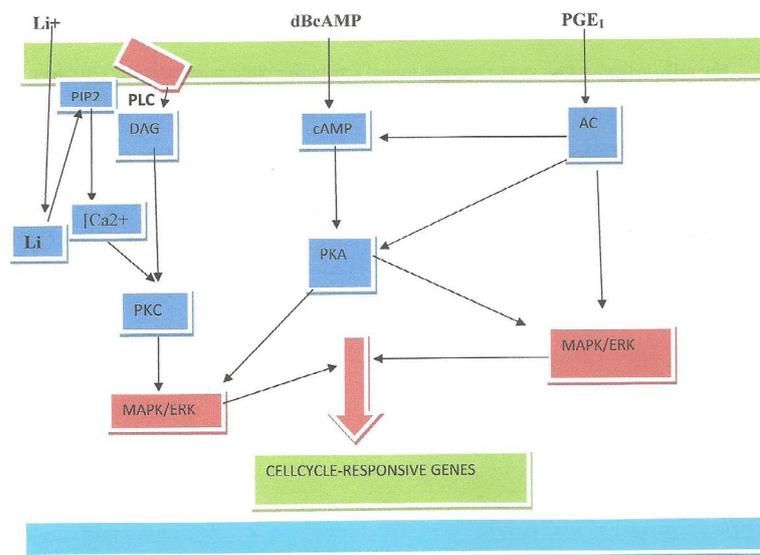
**Figure 3.** Photographs of clonal growth assay culture dishes showing the enhancing effect of PGE1 in combination with EGF and Insulin cultured in the presence of 1X level of amino acid (left panel) and in presence of 3X amino acids (right panel) on the clonal growth of normal human keratinocytes. A) control culture dishes supplemented with I<sup>+</sup>/E<sup>+</sup>; b) experimental supplemented with I<sup>+</sup>/E<sup>+</sup>/PGE1. All dishes were fixed with 5% glutaraldehyde and stained with 0.2% Crystal Violet stain. The dishes were photographed (1X, magnification).



**Figure 4.** Photographs of clonal growth assay culture dishes showing the enhancing effect of Lithium ions (Li<sup>+</sup>) in combination with EGF and IGF-1 when cultured in the presence of 1X amino acid (left panel), and no enhancement when cultured in presence of 3X amino acids (right panel) on the clonal growth of normal human keratinocytes. A) control culture dishes supplemented with I<sup>+</sup> and E<sup>+</sup> only; B) experimental culture dishes supplemented with I<sup>+</sup> and E<sup>+</sup> plus PGE1. All dishes were fixed with 5% glutaraldehyde and stained with 0.2% Crystal Violet stain. The dishes were photographed (1X, magnification).

Lithium ions presented as a salt, lithium chloride, is readily soluble in the aqueous phase of the medium and can be added aseptically as a 1:100 fold dilution and as the terminal component from a 1M LiCl concentrated stock solution to give a final concentration of 10 mM. Likewise, a 100 mM stock solution of dibutyl-cAMP dissolved in alcohol can be diluted 1:1000 aseptically to yield a final concentration of 0.1 mM and added as the terminal component to the final basal nutrient medium. Finally, 1 mg/mL stock solution of prostaglandin E<sub>1</sub> can be diluted 1:1000 aseptically to yield a final concentration of 10 μg/mL in the culture medium as the terminal component of the medium.

Lithium ions qualify per se as an independent growth factor. The example of MCDB153 as a SFM is one of several SFM medium composition that could be improved by addition of one or more and combinations of the named non-protein growth factors to commercially available serum-free culture media designed for keratinocytes. Earlier work, reported that Lithium Carbonate stimulated the proliferation of keratinocytes in *in vitro* normal human skin explants [21]. The effect occurred in a media supplemented with 10% fetal calf serum raising doubts about confounding effects of serum, and the evidence for keratinocyte proliferation was indirect microscope observations of cell crowding. In another tissue, the serum-free growth of human mammary epithelial cells is supported by addition of pituitary extracts [22]. Unfortunately, this addition defeats the purpose of having only defined growth promoting ingredients. Other non human cell studies have shown that Lithium can interfere with morphological movements and gene expression in the slime mold *Dictyostelium discoideum* [23], and embryological processes such as gastrulation in sea urchin [24], and amphibian embryos [25]. A possible mechanism by which Lithium ions interfere with cellular movements is by stabilizing cytoskeleton F-actin fibers [26,27]. More relevant to effects on cell growth is a report that Li<sup>+</sup> ions cause morphological alterations in various mammalian cell lines [28] at concentrations above 4 mM. The mechanism of action of Lithium ions is largely unknown but it can affect the intracellular phosphoinositide signaling pathway through depletion of the intermediate, inositol triphosphate (see Figure 5). In addition, when added to the culture medium of the ciliate, *Blepharisma*, it had an inhibitory effect on that cells photoresponses [29]. Lithium ions at 15 mM when added to culture medium of the protozoan parasite, *Herpetomonas*, stimulated its growth [30].



**Figure 5.** A hypothetical model of the mode of action of Lithium ions(Li<sup>+</sup>), dibutyl 3'-5' cyclic adenosine monophosphate (dBcAMP), and prostaglandin E<sub>1</sub> (PGE<sub>1</sub>) on activation of cell proliferation. Symbola: plasma-membrane ; phospholipase C, PLC, ; adenyl cyclase, ; phosphokinase A ; phosphokinase C ; mitogen-activated protein kinase, ; diacylglycerol, DAG; phosphoinositol-diphosphate, PIP<sub>2</sub>; calcium ion, [Ca<sup>2+</sup>].

Lithium ions at 20 mM concentration inhibited glucose synthesis in the rat liver [31]. Most relevant to the present report that Lithium ions at a concentration of 2-20 mM had a stimulatory effect on insulin induced uptake of α-aminobutyric acid, synthesis of DNA and RNA and cell multiplication of mouse mammary gland explants cultured in a chemically-defined synthetic medium.

Cyclic AMP is synthesized from ATP by adenylcyclase, a plasma membrane enzyme, which is ordinarily activated by hormone ligand occupation of specific cell surface hormone receptors like the ligand, insulin. cAMP is an intracellular second messenger that is involved in a signaling cascade, which acts through a cAMP-dependent protein kinase (PKA) that phosphorylates nuclear DNA binding proteins, thereby turning on select DNA sequences for gene expression. cAMP also activates calcium channels, providing a minor pathway by which cAMP causes a release of growth hormone. In non-human systems, cAMP is a chemoattractant for slime mold cells. Like Lithium ions, cAMP through cAMP-dependent PKA regulates actin organization and cell motility [13].

We also report that dBcAMP enhances keratinocyte proliferation based on dramatic increase in number and size of colonies after 10 days of clonal growth. Earlier workers, found a modest increment in population doubling of less than 1 doubling in 7 days of batch culture growth in SFM [32]. Contrary to these findings, another author found that dBcAMP had no direct effect on keratinocyte proliferation [33]. Their 5-Brdur DNA synthesis detection method may have missed a minor stimulation in cell proliferation.

Previously, PGE<sub>1</sub> and PGI and its analogues were shown to stimulate keratinocyte proliferation when treated with conditioned media containing cytokines released into the media by fibroblasts [34,35]. By contrast, we found that PGE<sub>1</sub> directly stimulates serum-free clonal growth in the presence of IGF-1 and in the total absence of any added protein growth factor.

In summary, we propose (see Figure 5) a hypothetical model of the mode of action of Lithium ions, diBcAMP, and PGE<sub>1</sub> as they impinge of the gene expression affecting the cell division cycle. We consider non-protein growth factors as first messengers or extracellular substances that include protein growth factors. As first messengers may not physically cross the phospholipid bilayer-cell membrane to initiate changes within the cell directly necessitating a second signal transduction mechanisms to transduce the first into second messengers, so that the extracellular signal may be propagated intracellularly and trigger proliferation. Second messengers triggered by Lithium ions are known parts of the phosphoinositide signalling pathway (A) which activates phosphoinositol kinase with the production of three second messengers, inositol triphosphate, diacylglycerol, and calcium. DBcAMP and PGE<sub>1</sub> act by activating the cAMP second messenger pathway (B and C). The cAMP second messenger signalling cascade involves downstream multi-cyclic kinases that link to mitogen-activated protein kinase (MAPK) and to transcription factors such as MYC that advance cell proliferation (see Figure 5 (A-C)).

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