A rare case of autoimmune pancytopenia (AIP) preceded by refractory anemia: possible involvement of cytomegalovirus (CMV) infection in pathogenesis

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Abstract
Autoimmune pancytopenia (AIP) is known to be caused by coexisting autoantibodies against three lineages of blood cells. We report a rare case of autoimmune pancytopenia (AIP) which was preceded by refractory anemia. An 85-year-old man with a 4 and half-year history of unexplained anemia presented in October 2016 because of remarkable pancytopenia and fever. Since direct and indirect anti-globulin tests as well as anti-platelet antibody were positive, Evans’ syndrome was initially suspected. However, the patient also had remarkable neutropenia. Additional examination for anti-neutrophil antibody showed the presence of anti-neutrophil IgG antibody specific for human neutrophil alloantigen (HNA)-1a and HNA-2. 

Case presentation
An 85-year-old man (49 kg/161cm) with a 4 and half-year history of unexplained anemia (red blood cell [RBC]: 3.31~3.91 × 10^12/L, hemoglobin [Hb]: 10.8~12.6g/dL) and hypertension since mid-fifties of age was referred to Imai Hospital from a family physician on October 25, 2016 because of oral cavity bleeding and remarkable pancytopenia. In 1949, Evans and Duane [1] first reported the association of acquired autoimmune hemolytic anemia (AIHA) with thrombocytopenia and/or leukopenia. In 1951, they reported 4 cases of AIHA associated with primary thrombocytopenic purpura, and suggested a possible antibody-mediated autoimmune mechanism as the cause [2].

Recently, we encountered a rare case of AIP preceded by refractory anemia in which cytomegalovirus (CMV) reactivation was demonstrated. In this paper, the importance of recognizing AIP in cases of Evans’ syndrome associated with leukocytopenia is emphasized.
During 3 days before admission, the patient was febrile with sore throat. On admission, he had a temperature of 38°C, and seemed pale and exhausted. There were several submucosal hemorrhages on the palate and hematomas on the lower lip and the left buccal mucosa. Multiple purpuras were noted over the surfaces of upper and lower extremities. Hepatosplenomegaly and superficial lymph node swellings were absent. His past history was somewhat complicated. He has been troubled with lumbar disc hernia since November 2015 and was previously treated with loxoprofen by a local orthopedic surgeon. In March 2016, acute renal failure associated with severe thrombocytopenia (30 × 10^9/L), possibly caused by loxoprofen administration, developed. Both disorders were successfully treated with the hemodialysis for a short period of time in other hospital (the exact cause of thrombocytopenia remains unknown). On September 6, 2016, the patient received a laminectomy of 4th and 5th lumbar vertebrae in an orthopedic division of another hospital without receiving any blood transfusions. The postoperative course was uneventful (Figure 1).

On admission (October 25, 2016), a complete blood cell count showed anemia (RBC count, 2.34 × 10^12/L; Hb, 7.3 g/dL; reticulocytes, 15.4%), leukocytopenia (white blood cell [WBC] count, 1.1 × 10^9/L with 40% neutrophils, 24% lymphocytes, 32% monocytes, 3% eosinophils and 1% basophils) and thrombocytopenia (platelet count, 1.0 × 10^9/L). Elevated levels of CRP (9.73 mg/dL; nr. 0–0.26 mg/dL), total bilirubin (1.57 mg/dL; nr. 0.2–1.2 mg/dL), indirect bilirubin (1.3 mg/dL; nr. 0.1–0.3 mg/dL), LDH (255 IU/L; nr. 106–211 IU/L), BUN (23.8 mg/dL; nr. 8–20 mg/dL), soluble-IL-2 receptors (s-IL-2R) (1,310 U/mL; nr. 124–466 U/mL), β2-microglobulin (3.3 mg/L; nr. 0.9–1.9 mg/L), ferritin (551 mg/dL; nr. 15–116 mg/dL), albumin (3.22 g/dL; nr. 3.9–4.9 g/dL) and creatinine (1.6 mg/dL; nr. 0.2–1.2 mg/dL). Direct and indirect anti-globulin tests and positive anti-platelet antibody were noted. Levels of anti-neutrophil, anti-macrophage, anti-epithelial, anti-thrombocyte, anti-rheumatoid factor, anti-erythrocyte, anti-platelet antibodies were confirmed. Direct and indirect anti-globulin tests and anti-platelet antibody were positive. Serum levels of creatinine, CEA, CA19-9, SLX, immunoglobulin (Ig) G/A/M, anti-nuclear antibody, anti-thyroglobulin antibody, anti-microsome antibody were positive. Serum levels of IgG, IgA, IgM antibodies against Parvovirus were not detected. A positron emission computed tomography scan, which was carried out 3 weeks after admission, showed no notable 18F-fluorodeoxyglucose uptake; neither lymphadenopathy nor splenomegaly was present (not shown). The laboratory test results showing hemolysis, positive direct/indirect antiglobulin tests and positive anti-platelet antibody led to a tentative diagnosis of Evans’ syndrome. Since the patient concomitantly had remarkable neutropenia, anti-neutrophil antibodies were later examined by using the serum stocked at initial presentation and examined by applying five cell-lineage immunofluorescence test (IFT) [12] and LABScreen MULTI (One Lambda, Inc.) assay [13]. As expected, the patient was found to have anti-neutrophil IgG antibodies specific for human neutrophil alloantigen (HNA)-1a and HNA-2 (Figure 2). This, a diagnosis of autoimmune pancytopenia (AIP) was later established.

After admission, administration of prednisolone (PSL) (40 mg/day) was immediately initiated for 3 days. Although the temperature

Figure 1. Clinical course before admission. The patient had slight anemia, and it appeared 4 and half years before admission. Acute renal failure (ARF) caused by loxoprofen administration developed in March 2016, but this was improved by hemodialysis (HD). The ARF was associated with remarkable thrombocytopenia (Plt: 3 × 10^9/L). Both disorders were improved by HD. On admission, the remarkable pancytopenia was seen.

Figure 2. (A) Five cell-lineage IFT showing positive IgG antibodies (asterisk) against neutrophils and platelets. However, the latter positivity was considered to be platelet-associated antibody (possibly anti-platelet IgG antibody) other than HNA, because HNA-1a and HNA-2 are not usually expressed on platelets. Whole peripheral blood samples were obtained from six healthy volunteers (sample 1–6) and mixed with the patient’s serum. Bound antibodies on 5 different blood cell lineages were demonstrated by using fluorescein isothiocyanate-conjugated goat anti-human immunoglobulin G (IgG) and analyzed by flow cytometry. The autologous sera of volunteers were used as negative controls. A positive cut-off was set at more than two times the average fluorescence of negative control of the healthy volunteers.

(B) LABScreen MULTI assay demonstrating anti-neutrophil IgG antibody specific for HNA-1a and HNA-2 (asterisk). The basis of the LABScreen MULTI is the use of color-coded microbeads coated with HNA peptides. Data acquisition and analysis were performed by the Lambda Assay Beads MULTI Analyte system using the LABScan 100 flow analyzer.
Takemori N (2017) A rare case of autoimmune pancytopenia (AIP) preceded by refractory anemia: possible involvement of cytomegalovirus (CMV) infection in pathogenesis

dropped rapidly by PSL administration, the pancytopenia was not improved. Then a high-dose methyl-prednisolone (m-PSL) therapy (1000 mg/day for 3 days with gradual reduction schedule), which was followed by reduced dose of m-PSL and then conventional dose of PSL therapy, was initiated. The steroid therapy was effective; depressed platelet count, neutropenia and elevated CRP level returned to normal 1 week after the initiation of the treatment. The indirect antiglobulin test returned to negative 1 month after the initiation of the steroid therapy, but the direct antiglobulin test remained positive. Anti-platelet antibody became negative about 10 days after admission. In this case, cyclosporine (200–400 mg/day) and metenolone (15 mg/day) were added to the steroid therapy 2 weeks later because of insufficient recovery from anemia (both drugs were later suspended because of ineffectiveness on anemia). With the reduction of the PSL dose, severe thrombocytopenia associated with positive anti-platelet antibody reappeared 2 months after admission, but this was improved by a semi-high-dose m-PSL (500 mg/day for 3 days) therapy (Figure 3). This event, a febrile episode and elevated levels of CRP, β2-microglobulin and s-IL-2R at initial presentation prompted us to investigate the possibility of specific infections by using the stocked serum. Consequently, the patient was found to have cytomegalovirus (CMV) infection; the presence of CMV-IgM antibody as well as CMV-DNA was demonstrated in the serum stocked at the reappearance of thrombocytopenia. Serum stocked at initial presentation was retrospectively examined for CMV-IgG and IgM. The results revealed positive CMV-IgG and negative CMV-IgM. Thus, it was considered that a reactivation of CMV has occurred during the hospital course. The reappeared anti-platelet antibody returned to negative after the recovery from thrombocytopenia (Table 1 and Figure 3).

In the present case, bone marrow (BM) examination could not be done before initiation of the treatment because of patient’s exhausted condition, severe anemia and severe thrombocytopenia. Later, the BM aspiration tests were carried out on three occasions during the treatment (November 18, 2106, December 13, 2016 and January 17, 2017). The first examination results revealed erythroid hyperplasia (M/E ratio, 0.5; nr 1.2–3.8) and myelodysplastic features in megakaryocytes and erythroblasts. Micro-megakaryocytes and mononuclear megakaryocytes were frequently seen, and bimucled and trinucleated erythroblasts were sporadically seen. Mature megakaryocytes showing platelet productions were absent. The second and third examinations revealed hypocellular marrows but normal in differential count; however, though very rare, bimucled and trinucleated erythroblasts were seen. Phagocytic findings were absent (Figure 4). Chromosome analyses consistently showed a normal karyotype. Thus, the patient was diagnosed as having refractory anemia (according to the FAB classification of the myelodysplastic syndrome [MDS] [14]), refractory cytopenia with unilineage dysplasia (according to the 2008 revision of the WHO classification [15]) or MDS with single lineage dysplasia (according to the 2016 revision of WHO classification [16]). The patient was discharged on April 22, 2017 and currently remains in good condition (Figure 3); however, CMV-IgM is persistently positive as of May 7, 2017.

**Discussion**

For the correct diagnosis of AIP, the demonstration of autoantibodies against three lineages of blood cells is required. The methods to detect autoantibodies against RBCs and platelets are widely used in the clinical field. However, the availability of methods to demonstrate anti-neutrophil antibodies is limited in the clinics. AIN is known as a common form of neutropenia in childhood and is characterized by chronic neutropenia presenting a low absolute neutrophil count of less than 1.5 × 10^9/L for more than 6 months [3–6]. This definition of neutropenia is applied also to neutropenia of AIP. Recently, Matsuyama, et al. [12] developed a flow cytometric (FCM) analysis system to detect anti-neutrophil antibodies (i.e., five cell-lineage IFT system). This system made it possible to simultaneously detect antibodies against 5 different individual leukocytes (neutrophils, monocytes, CD4+ T cells, CD20+ B-cells and platelets). Using this system together with LABScreen microbeads test (i.e., LABScreen MULT1 assay system), we were able to demonstrate anti-neutrophil IgG antibody specific for HNA-1a and HNA-2. Thus, it was proved that the patient had autoantibodies for

| Table 1. Levels of CMV-IgM/IgG/DNA, autoantibodies, hematologic data, CRP and s-IL-2R during clinical course. |
|---|---|---|---|---|---|---|
| Date | 10/25/2016 | 12/2 | 12/24 | 2/7/2017 | 3/6 | 4/14 |
| CMV-IgM (+, index) | < 0.8: (―) | 5.01: (+) | 8.9: (+) | 9.2: (+) | 7.83: (+) | 6.77: (+) |
| CMV-IgG (< 2.0, EIA) | 14.3: (+) | 19200: (+) |
| CMV-DNA (< 100 C/mL) | (―/―) |
| Platelet antibody | (+) | (―) | (―) | (―) | (―) | (―) |
| Direct/indirect AGT | (+/+) | (+/+) | (+/+) | (―/―) | (―/―) | (―/―) |
| Anti-neutAb | (+) |
| Platelet (<10^10/L) | 1.0 | 24.7 | 2.2 | 34.8 | 34.8 | 23.4 |
| Hb (g/dL) | 7.3 | 7.6 | 9.2 | 9.9 | 11.4 | 10.9 |
| Neutrophil (>10^9/L) | 0.42 | 4.71 | 5.39 | 6.80 | 5.92 | 3.70 |
| CRP (mg/dL) | 9.73 | 0.20 | 0.02 | 0.05 | 0.05 | 0.02 |
| s-IL-2R (U/L) | 1310 | 1200 | 758 | 591 | 844 | 484 |

Abbreviations: AGT, antiglobulin test; Anti-neutAb, anti-neutrophil antibody; EIA, enzyme immune assay, Hb, hemoglobin, CRP, C reactive protein; s-IL-2R, soluble IL-2 receptors.
Takemori N (2017) A rare case of autoimmune pancytopenia (AIP) preceded by refractory anemia: possible involvement of cytomegalovirus (CMV) infection in pathogenesis

doi: 10.15761/HMO.1000128

Hematol Med Oncol, 2017

Figure 4. BM smear (May–Grünwald–Giemsa stain, under×40 magnification objective). (A) A micromegakaryocyte (arrow) seen in the BM obtained on November 18, 2016. (B) A small mononuclear megakaryocyte without platelet production (arrow) seen in the bone marrow obtained on November 18, 2016. Normal mature megakaryocytes were absent in the BM (not shown). (C) A binucleated erythroblast seen in the BM obtained on November 18, 2016. (D) A trinucleated erythroblast (arrow) seen in the BM obtained December 13, 2016.

Three lineages of blood cells, and the diagnosis of AIP was established. Much less is known about idiopathic AIP. Wiesneth, et al. [7] reported 2 cases of idiopathic AIP in 55 cases of unexplained neutropenia, Bux, et al. [5] reported 3 cases of idiopathic AIP in 143 cases of AIN, and Martino, et al. [8] reported 2 cases of idiopathic AIP in 20 cases of AIN, and recently, Mitani, et al. [11] reported 1 case of idiopathic AIP. Thus, 8 cases of idiopathic AIP have been documented in the literature. To establish the diagnosis of idiopathic AIP, it is necessary to completely rule out collagen disease, rheumatoid arthritis, lymphoproliferative disorder, cancer and specific infection causing immune dysfunctions. In the present case, CMV infection was confirmed during the course of illness. In addition, refractory anemia preceded before developing AIP. Thus, this case might represent an acquired AIP due to CMV infection/reactivation.

Human CMV is a β human herpesvirus and can cause life-threatening infections in immunocompromised patients. It is known that CMV infection can cause thrombocytopenia in immunocompetent adults as well as children [17-25]. In addition, CMV infection is known to trigger an autoimmune process against platelets even though the infection itself is transient and seemingly neither atypical nor severe in nature [19]. CMV infection is also known to cause autoimmune hemolytic anemia [26,27]. In this connection, van Sproos and Breed [28] reported a case of severe symptomatic thrombocytopenia combined with hemolytic anemia which was caused by CMV infection. Koukoulaki, et al. [29] reported a rare case of a previously healthy woman who developed irreversible pancytopenia secondary to CMV infection in which no autoantibody was detected. To our knowledge, there is no report documenting AIP caused by CMV infection/reactivation.

In the present case, positive CMV-IgG at initial presentation means that the patient was previously infected quiescently by CMV. The reappearance of thrombocytopenia during the hospital course can be considered to be caused by reactivation of CMV based on the finding of CMV-DNA and CMV-IgM during thrombocytopenia. Since anti-platelet antibody reappeared during thrombocytopenia but returned to negative after improvement of thrombocytopenia, it clearly showed that autoimmune mechanism is involved in developing thrombocytopenia. In the present case, CMV-IgM was negative at initial presentation; however, it should be noted that it takes about 1 week for CMV-IgM to appear in the serum after the beginning of symptoms. In this case, blood sampling was done only 3 days after the beginning of symptoms. Thus, it is conceivable that seroconversion did not yet occurred at initial presentation. Rather, it seems plausible that CMV reactivation had actually occurred at the time of serum sampling. The laboratory data at initial presentation which showed acute inflammatory reactions (e.g., elevated levels of CRP, β2 microglobulin and s-IL-2R) and clinical symptoms such as fever and sore throat suggested recent CMV reactivation. It is well known that s-IL-2R will increase via immune activation in cases of viral infection. Thus, CMV reactivation might have also occurred at initial presentation and possibly caused AIP. In the present case, CMV-IgM is still persisting. It is known that in some cases, CMV-IgM may persist for up to 2 years.

Two possible mechanisms have been proposed for the pathogenesis of cytopenia(s) caused by CMV infection: a direct cytopathic effect of CMV on hematopoietic cells and an indirect immune-mediated effect [18]. In the former, several in vitro studies have shown that certain strains of CMV can infect hematopoietic progenitor cells and substantially inhibit hematopoiesis [30,31,32]. In addition, it has been suggested that CMV infection results in downregulation of G-CSF and hematopoietic suppression [31]. Singh and Ruscetti [32] reported that BM cells infected with CMV became totally insensitive to the stimulatory effects of both GM-CSF and G-CSF [31]. Concerning the indirect immune-mediated mechanism, molecular mimicry of CMV has been implicated in the onset of autoimmunity [33]. Viral peptides that share homology with hosts-peptides could initiate an immune response both against itself and the virus. Additionally, during viral cell lysis, intracellular proteins, the secured host antigens not previously recognized, are exposed to circulating antigenpresenting cells that activate autoreactive lymphocytes. In addition, the possibility that hemophagocytic syndrome triggered by CMV can result in the destruction of hematopoietic cells, is also indicated [34]. However, no evidence of hemophagocytosis was found in the bone marrow in our case. Since neutropenia and hemolytic anemia were not exaggerated during the thrombocytopenia reappearance, it seems that megakaryocytic lineage might be more sensitive to CMV infection/reactivation than myeloid and erythroid lineages.

In the present case, refractory anemia preceded the development of AIP. It is known that autoimmune paraneoplastic syndromes are commonly encountered in patients with MDS. As many as 10% of MDS patients may experience various autoimmune syndromes [35]. Hemolytic anemia and ITP are known to be seen, though rare, in patients with MDS [35-37]. However, the direct relationship between refractory anemia and AIP seems improbable based upon the patient’s clinical course. Rather, it seems probable that AIP simply happened to be complicated by refractory anemia in this case.

Concerning elevated level of IgE at initial presentation, it seems to reflect patient’s allergic disposition. This might have been related to previous episode of acute renal failure and thrombocytopenia due toloxopran administration. However, this possibility has remained in the realm of surmise due to the lack of clinical data. Furthermore, the relationship between this episode and CMV infection remains unknown.

The distinction between direct and indirect effect of CMV is
important because it may have very important therapeutic implications: an immune-mediated effect would be expected to respond to steroids, but a cytotoxic effect would not; the direct mechanism presupposes a good response to specific anti-CMV treatments [25]. However, literature review suggested that corticosteroids are also effective in approximately one third of cases [20]. Some physicians believe that treatment with steroids should be avoided in patients with CMV-induced thrombocytopenia because immunosuppressive treatment may cause the exacerbation of primary CMV infection and trigger further decrease in platelet count [24]. If CMV-viremia is demonstrated, viral eradication seems to have a positive influence on the resolution of thrombocytopenia [23]. Treatments with ganciclovir or valganciclovir is currently recommended as first line treatment for immunocompromised adults with severe CMV disease, and few studies have evaluated the use of these anti-CMV drugs for the treatment in immunocompetent adults [38]. The use of foscarnet, which does not share the myelosuppressive effects of ganciclovir and valganciclovir, seems to be a reasonable first line anti-CMV agent in cases of cytopenia [23]. Splenectomy [21,23,24] and intravenous immunoglobulin [19,22,24,], useful in patients affected by idiopathic thrombocytopenic purpura, are relatively ineffectual or refractory for CMV-induced thrombocytopenia. However, it is known that splenectomy regain their efficacy once the primary CMV infection is controlled with anti-CMV agents [23,24]. In the present case, anti-viral agents were not used because of remarkable effects of steroid therapy, and because CMV infection/reactivation was not presupposed and retrospectively confirmed after the initiation of steroid therapy. If relapse were to occur in the present case, anti-CMV treatment should be considered. In cases of unexplained acute pancytopenia associated with clinical manifestations of an infectious disease, testing for CMV should be required for better therapeutic consequences.

Conclusions

A rare case of acquired AIP associated refractory anemia is documented. In particular, the recognition of AIP should be requisite to some cases of Evans’ syndrome associated with leukocytopenia. Special consideration should be given to the possible involvement of CMV infection in pathogenesis of immune cytopenia(s) or AIP. We emphasize the importance of testing for anti-neutrophil antibody in cases of pancytopenia and AIP. Insufficient recovery from anemia in the present case seemed to be related to coexisting refractory anemia.

Acknowledgements

The authors would like to thank Nobuki Matsuyama (medical anti-neutrophil antibody and providing the data), and Prof. Hong-Kean Ooi (Department of Veterinary Medicine, Azabu University) for helpful reviewing and preparing the manuscript.

Consent

Written informed consent was obtained from the patient for publication of this case report and accompanying images.

Competing interests

The authors declare no competing interests.

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