

Table S1. Detailed overview on the parameters used for the systematic assessment by both diagnostic techniques

Documented parameters by cytomorphology	Documented parameters by Histopathology	
Granulopoiesis		
<ul style="list-style-type: none">—Toxic/Dysplastic granulation—Hypogranularity of cytoplasm—Pseudopelger	<ul style="list-style-type: none">—Left-shifted—Maturation disorder	
Erythropoiesis		
<ul style="list-style-type: none">—Left-shift—Erythroblasts with vacuolated cytoplasm—Megaloblastoid—Irregular nuclei—Nuclear lobulation—Bi-/Multinucleated cells—Fragmentation of nuclei—Internuclear bridges—Pyknosis of nuclei—Erythrons—Gigantism	<ul style="list-style-type: none">—Left-shift—Maturation disorder—Megaloblastoid—Irregular nuclei—Bi-/Multinucleated cells	
Megakaryopoiesis		
<ul style="list-style-type: none">—Microkaryocytes—Multiple separate nuclei—Poor granulation of cytoplasm—Bizarre nuclear shape—Giant forms	<ul style="list-style-type: none">—Microkaryocytes—Multiple separate nuclei—Maturation disorder—Hyperchromatic nuclei—Hyperlobated nuclei—Monolobated nuclei—Hypolobated nuclei—Cluster of megakaryocytes—Giant forms—Abnormal localization	
Special Stains / Immunohistochemistry		
<ul style="list-style-type: none">—Non-specific Esterase—Iron (Berliner Blau)—Myeloperoxidase	<ul style="list-style-type: none">—CD3—CD20—CD34—CD42b—CD68—CD71	<ul style="list-style-type: none">—CD117—CD138—Iron (Berliner Blau)—Myeloperoxidase—Gomori

Table S2A. Dysplasia in in the different hematopoietic lineages detected by cytomorphology and histopathology

	Number of cases (%)		
	Granulopoiesis	Erythroipoiesis	Megakaryopoiesis
Cytomorphology			
Evaluable cases	82	82	65
Dysplasia ≥ 10%	75/82 (91.5%)	82/82 (100.0%)	53/65 (81.5%)
Dysplasia ≥ 10% <30%	4/82 (4.9%)	1/82 (1.2%)	9/65 (13.8%)
Dysplasia ≥ 30%	35/82 (42.7%)	31/82 (37.8%)	20/65 (30.8%)
Dysplasia ≥ 60%	36/82 (43.9%)	50/82 (61.0%)	24/65 (36.9%)
Histopathology			
Evaluable cases	86	79	82
Maturation disorder	71/86 (82.6%)	77/79 (97.5%)	80/82 (97.6%)
Mild maturation disorder	55/86 (64.0%)	49/79 (62.0%)	40/82 (48.8%)
Severe maturation disorder	16/86 (18.6%)	28/79 (35.4%)	40/82 (48.8%)
Correlation of both methods			
Results of χ^2 -Test	$\chi^2(1)=1.176$, p=0.278	Not evaluable	$\chi^2(1)=1.587$, p=0.208

Table S2B. Overall cellularity and cellularity of the distinct hematopoietic lineages as documented by cytomorphology and histopathology

Parameter	Cytomorphology N (%)	Histopathology N (%)
Overall cellularity	83 (100.0%)	90 (100.0%)
Normal (age-adjusted)	10 (12.0%)	24 (26.7%)
Increased	66 (79.5%)	58 (64.4%)
Reduced	7 (8.4%)	8 (8.9%)
Correlation of both methods	$\chi^2(4)=7.942$, $p=0.094$	
Granulopoiesis	84 (100.0%)	87 (100.0%)
Normal (age-adjusted)	19 (22.6%)	24 (27.6%)
Increased	43 (51.2%)	39 (44.8%)
Reduced	22 (26.2%)	24 (27.6%)
Correlation of both methods	$\chi^2(4)=13.252$, $p=0.010$	
Erythropoiesis	85 (100.0%)	88 (100.0%)
Normal (age-adjusted)	29 (34.1%)	20 (22.7%)
Increased	30 (35.3%)	28 (31.8%)
Reduced	26 (30.6%)	40 (45.5%)
Correlation of both methods	$\chi^2(4)=26.764$, $p<0.001$	
Megakaryopoiesis	82 (100.0%)	86 (100.0%)
Normal (age-adjusted)	23 (28.0 %)	24 (27.9%)
Increased	29 (35.4%)	37 (43.0%)
Reduced	30 (36.6%)	25 (29.1%)
Correlation of both methods	$\chi^2(4)=36.180$, $p<0.001$	

Table S3. Results of molecular analysis for selected genes are given either by hotspots analysis and/or by NGS panel sequencing

Detected molecular mutations	
<i>ACD</i>	N=1/27
<i>ASXL1</i>	N=1/27
<i>CEBPA</i>	N=1/27
<i>DNMT3A</i>	N=2/27
<i>ETV6</i>	N=1/27
<i>EVII</i>	N=1/27
<i>FLT3</i>	N=2/27
<i>JAK2V617F</i>	N=1/27
<i>MECOM</i>	N=1/27
<i>MLL (=KMT2A)</i>	N=3/27
<i>NPM1</i>	N=1/27
<i>NRAS</i>	N=3/27
<i>RTEL1</i>	N=1/27
<i>RUNX1</i>	N=5/27
<i>SRSF2</i>	N=1/27
<i>TERC</i>	N=1/27
<i>U2AF1</i>	N=1/27
<i>TP53</i> ¹ analysed	N=8
positive	N=2

Analyses were performed by different laboratories on a physician-driven basis. Molecular analyses were available in 39 patients from the cohort (in 43.4% of all patients). Analysis of *TP53* mutations was performed in 8 patients, ¹*TP53* analysis only in MDS

Table S4. Detailed information of 18 cases with a discrepant final diagnosis by CM and HP

Age ¹ , Gender ²	Cytomorphology	Histopathology
69, m	RCMD	reactive
See Figure 1a	Slight hypercellularity Amount of blasts 4% Distinct dysplastic GP Increased, dysplastic EP Dysplastic MP	60% cellularity CD34+ blasts <5% Reactive GP No dysplasia in EP and MP
	NKT ³ , NGS Panel for AML/MDS ⁴ : no mutation	
79, f	RCMD	reactive
	Hypercellularity Amount of blasts 2% Dysplastic GP Dysplastic and left-shifted EP Decreased MP	40% cellularity Only slightly dysplastic GP, EP, MP Distinct reactive changes
	NKT ³ , no molecular analysis performed, pancytopenia	
75, m	RAEB-1	RCMD

See Figure 1b	Hypercellularity Amount of blasts 7% Increased, dysplastic, left-shifted GP Dysplastic, slightly left-shifted EP Increased, dysplastic, left-shifted MP	90% cellularity CD34+ blasts <5% CD68+ <5% Distinct dysplastic GP Macrocytic EP Normal MP
	NKT ³ , no molecular analysis performed, pancytopenia	
74, m	RAEB-1	RCMD
	Hypercellularity Amount of blasts 6% Increased, slightly dysplastic GP Decreased, dysplastic, left-shifted EP Increased, slightly left-shifted MP	70% cellularity CD34+ blasts <5% Slightly increased GP Distinct decreased, dysplastic EP Slightly increased, dysplastic MP
	NKT ³ , NGS Panel for AML/MDS ⁴ : no mutation	
74, f	RAEB-1	RCMD
	Hypercellularity Amount of blasts 5% Distinct increased, dysplastic GP Increased, dysplastic EP, MP	90% cellularity CD34+ blasts <5% Distinct increased, slightly dysplastic, left-shifted GP Decreased, macrocytic EP Dysplastic MP
	NKT ³ , PCR negative for: <i>AML1-ETO</i> , <i>CBFβ-MYH11</i> , <i>FLT3-ITD</i> , <i>PML-RARα</i> , <i>EVI</i> , Sanger sequencing negative for: <i>CEBPA</i> , <i>NPM1</i> ; pancytopenia	
73, m	RAEB-1	RCMD
	Hypercellularity Amount of blasts 5% Decreased, dysplastic GP Increased, dysplastic EP, MP	50% cellularity CD34+ blasts slightly <5% CD117+ 5% Dysplastic, left-shifted GP, EP Increased, distinct dysplastic MP
	NKT ³ , no molecular analysis performed, pancytopenia	
74, f	RAEB-1	RCMD
	Normocellularity Amount of blasts 6% Increased, dysplastic GP, EP, MP	Insufficient material quality 50% cellularity Amount of blasts <5% Increased, slightly left-shifted GP Decreased, slightly dysplastic EP Slightly increased MP
	del(7q), <i>NRAS</i> mutation (Sanger sequencing)	
67, f	RAEB-1	s-AML
	Hypercellularity Amount of blasts 5% Decreased, dysplastic GP Increased, dysplastic EP Limited quality of MP	98% cellularity CD34+ blasts 20% CD68/CD117 blasts partial positivity Increased, left-shifted GP Decreased EP, MP
	del(20q), clinically driven molecular analysis: no mutation, pancytopenia	
82, f	RAEB-1	suspected MDS
	Hypercellularity Amount of blasts 5% Dysplastic GP Increased, dysplastic EP Slightly dysplastic MP	60% cellularity CD34+ blasts slightly <5% Reactive GP Increased EP, MP
	No cytogenetics and no molecular analysis performed, pancytopenia	
77, m	RAEB-2	s-AML
	Insufficient material quality Hypercellularity Amount of blasts 16% Increased, dysplastic GP Dysplastic EP Reduced, dysplastic MP Monocytes increased	80% cellularity CD34+ blasts 30% CD68 blasts partial positivity Increased, dysplastic, left-shifted GP Decreased EP, MP
	NKT ³ , clinically driven molecular analysis: no mutation, pancytopenia	
69, f	CMML-1	suspected MDS
	Hypercellularity Amount of blasts 3% Increased, dysplastic GP Dysplastic EP, MP Increased Monocytes (EST not available)	70% cellularity CD34+ blasts <5% CD117+ blasts <5% CD68 physiological Increased GP Slightly dysplastic EP, MP
	NKT ³ , <i>BCR-ABL1</i> , <i>JAK2V617F</i> , <i>CALR</i> , <i>MPL</i> negative, Lc ⁵ 29.0 G/L, monocytes 6.0 G/L	
76, f	CMML-2	RAEB-2

	Hypercellularity Amount of blasts 16% Increased, reactive GP Dysplastic EP, MP Increased Monocytes (EST not available)	Insufficient material quality 70% cellularity CD34+ blasts 15%, focal up to 20% CD68 not available Increased GP Decreased, macrocytic EP Dysplastic, distinct left-shifted MP
	Complex KYT ⁶ , <i>BCR-ABL</i> and <i>JAK2V617F</i> negative, bicytopenia	
70, f	CMML-2	suspected MDS
See Figure 1c	Hypercellularity Amount of blasts 11% Increased, dysplastic GP Dysplastic EP Increased MP Monocytes increased (EST not available)	55% cellularity CD34+ blasts <5% CD68 without abnormality Increased, dysplastic, slightly left-shifted GP Normal EP Slightly dysplastic MP
	NKT ³ , <i>BCR-ABL1</i> and <i>JAK2V617F</i> negative	
59, m	CMML-2	RAEB-2
	Hypercellularity Amount of blasts 18% Increased, dysplastic, left-shifted GP Dysplastic EP, MP EST positivity 20% POX defect	55% cellularity CD34+ blasts 15% Increased, dysplastic, left-shifted GP Dysplastic EP, MP
	del(7q), <i>RUNX1</i> mutation (Sanger sequencing), bicytopenia	
71, m	s-AML	CMML-1
	Hypercellularity Amount of blasts 30% Increased, dysplastic GP Reduced, dysplastic EP Reduced MP EST positivity 80% Monocytes increased (EST not available)	Insufficient material quality 90% cellularity CD34+ blasts <5% CD117+ >5% CD68 blasts partial positivity Increased GP Decreased, slightly left-shifted EP Increased, dysplastic MP
	<i>der(19)</i> , +8, no molecular analysis performed	
76, f	RARS-T	RAEB-1
	Hypercellularity Amount of blasts 3% Ringsideroblasts >15% Dysplastic GP Dysplastic, left-shifted EP Increased, dysplastic MP	60% cellularity CD34+ blasts 5%, focal 9% Normal maturation of GP Dysplastic, slightly left-shifted EP Increased, dysplastic MP
	NKT ³ , PCR negative for: <i>JAK2V617F</i> , <i>CALR</i> , <i>MPL</i> , plt ⁷ 562 G/L	
78, f	suspected MDS	RAEB-1
	Insufficient material quality Evidence of blasts Existing GP and EP Absent MP	60% cellularity CD34+ blasts 5% Dysplastic GP Decreased, distinct dysplastic EP Increased, distinct dysplastic MP
	Complex KT ⁶ , <i>TP53</i> mutation (Sanger sequencing), pancytopenia	
69, m	Suspected MDS	s-AML
	Insufficient material quality Amount of blasts cannot be evaluated Dysplastic GP	75% cellularity Amount of blasts 25% CD34+ blasts 6% CD117+ <5% Increased GP Increased, dysplastic EP Increased, dysplastic, left-shifted MP
	NKT ³ , clinically driven molecular analysis: no mutation	

¹Age at first diagnosis; ²f=female, m=male; ³NKT=normal karyotype; ⁴AML/MDS mutation panel: *ASXL1*, *CBL*, *CEBPA*, *DNMT3A*, *ETV6*, *EZH2*, *GATA2*, *IDH1*, *IDH2*, *JAK2*, *KRAS*, *NRAS*, *RUNX1*, *SETBP1*, *SF3B1*, *SRSF2*, *TET2*, *TP53*, *U2AF*, *ZRSR2*; ⁵Lc=leukocytes; ⁶KT=karyotype; ⁷Plt=platelets